

## Minor Triterpenoids from the Mediterranean Sponge, *Raspaciona aculeata*

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MINOR TRITERPENOIDS FROM THE MEDITERRANEAN  
SPONGE, *RASPACIONA ACULEATA*

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ABSTRACT.—Eight new minor triterpenoids have been characterized from a lipid-soluble extract of *Raspaciona aculeata*. All are based on the same carbon skeleton as raspacionin [1]. The full assignment of all  $^1\text{H}$ - and  $^{13}\text{C}$ -nmr resonances has led to the rationalization of the effects of certain substituents on the chemical shifts of atoms in the perhydrobenzoxepine system.

Raspacionin [1] and raspacionins A [2], and B [3] are three terpenoids recently found in the encrusting red sponge, *Raspaciona aculeata* Johnston (family Raspailiidae) (1–3). Their structures have been elucidated by X-ray analysis and by extensive nmr investigations. Their absolute stereochemistry has been suggested either by applying high-resolution  $^1\text{H}$  nmr (2,4) to the Mosher method (5,6), as successfully proposed for siphonolol A [4] by Kakisawa's group (7–10), or by comparison of cd spectra (3).

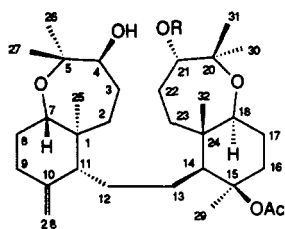
In this communication we now report the structural characterization of a further eight [5–12] minor components of *Raspaciona aculeata*, all based on the same triterpenoid skeleton as raspacionin [1]. The structures of the new raspacionins display different functionalizations at carbons 4, 10, 15, and 21. The full nmr characterization of 5–12 has revealed that some shifts of the substituents are diagnostic for both ring location and stereochemical orientation.

The only previous triterpenoids related to raspacionins, besides those from *Raspaciona aculeata*, have been those from the sponge *Siphonocalina siphonella* (11–15) and, more recently, from the South African purple-brown fan sponge *Axinella weltneri* (16).

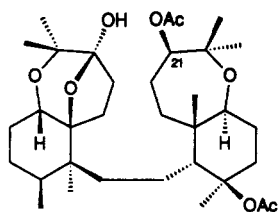
## RESULTS AND DISCUSSION

The lipid-soluble extract from *Raspaciona aculeata* was submitted to the usual work-up (2,3) yielding, along with 1 and a mixture of 2 and 3, a more polar fraction ( $R_f$  0.27, petroleum ether-Et<sub>2</sub>O, 3:7) that was purified by hplc (Spherisorb Silica column, *n*-hexane-EtOAc, 7:3) giving seven main fractions which, after further hplc purification, yielded eight pure isolates [5–12]. The structural characterization of the products is reported herein, starting from compound 5, which was most closely related to raspacionin [1], and then reporting the other metabolites according to their chemical affinity with 1.

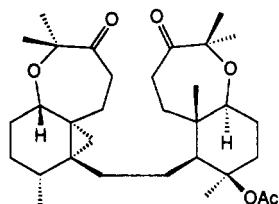
21-Deacetyl-raspacionin [5] is an optically active compound with elemental composition C<sub>32</sub>H<sub>54</sub>O<sub>6</sub> determined by hreims on the fragment ion peak at  $m/z$  474.3698 [ $\text{M}-\text{HOAc}$ ]<sup>+</sup> (C<sub>30</sub>H<sub>50</sub>O<sub>4</sub> requires 474.3709). The  $^1\text{H}$ -nmr spectrum of 5 was almost identical to that of raspacionin [1]; diagnostic differences were the chemical shift of H-21 ( $\delta$  3.82;  $\delta$  4.97 for 1) and the absence of the acetyl methyl singlet at  $\delta$  2.17. All  $^1\text{H}$ - and  $^{13}\text{C}$ -nmr resonances (Tables 1 and 2) were assigned by 1D and 2D nmr experiments (DEPT,  $^1\text{H}$ - $^{13}\text{C}$  HETCOR,  $^1\text{H}$ - $^1\text{H}$  COSY). By comparing the nmr data of 5 with those of raspacionin [1], it was observed that the absence of an acetyl group at C-21 induces, besides the expected shifts for the atoms near C-21, two  $^1\text{H}$ -nmr downfield shifts for H-14 ( $\delta$  0.83;  $\delta$  0.74 for 1) and H-18 ( $\delta$  3.55;  $\delta$  3.41 for 1). Methanolysis of 1 yielded a compound identical in all aspects to 5.



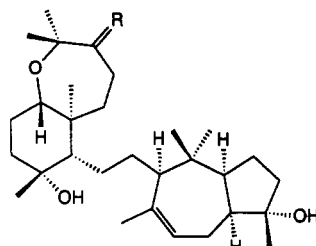
1 R=Ac  
5 R=H



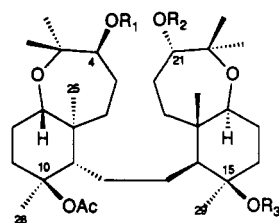
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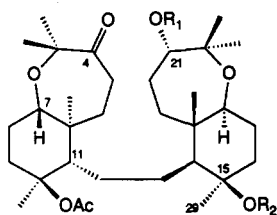
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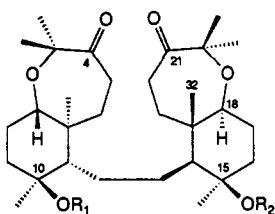
4 R=β-OH, H  
13 R=O



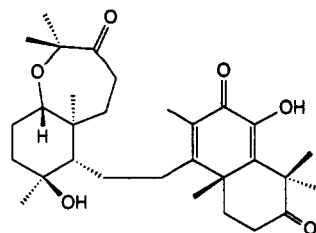
6 R<sub>1</sub>=H; R<sub>2</sub>=H; R<sub>3</sub>=Ac  
10 R<sub>1</sub>=Ac; R<sub>2</sub>=Ac; R<sub>3</sub>=H



7 R<sub>1</sub>=H; R<sub>2</sub>=Ac  
8 R<sub>1</sub>=H; R<sub>2</sub>=H  
9 R<sub>1</sub>=Ac; R<sub>2</sub>=H



11 R<sub>1</sub>=Ac; R<sub>2</sub>=H  
12 R<sub>1</sub>=H; R<sub>2</sub>=Ac



14

10-Acetoxy-21-deacetyl-28-hydroraspacionin [**6**], exhibited a molecular formula of  $C_{34}H_{58}O_8$ , deduced from hreims at  $m/z$  474.3684 (main mass fragment peak,  $[M-2HOAc]^+$ ;  $C_{30}H_{50}O_4$  requires 474.3709). The  $^1H$ -nmr spectrum of **6** was differentiated from that of **1** by the absence of the exomethylene protons and by the presence of two singlet methyls at  $\delta$  1.89 (OCOCH<sub>3</sub>-10) and  $\delta$  1.48 (H<sub>3</sub>-28), consistent with a different substitution pattern at carbons 10 and 28 in **6**. The  $^{13}C$ -nmr resonance of C-28 at  $\delta$  19.71 supported an axial orientation of this methyl functionality. The stereochemistry at C-10 was differentiated from that at C-15 by a series of diagnostic shifts. Thus, the equatorial orientation of the acetoxy group at C-10 induced  $^1H$ -nmr upfield shifts for the equatorial H-9 ( $\delta$  2.63; H-16 eq  $\delta$  2.78) and for H<sub>3</sub>-25 ( $\delta$  0.83; H<sub>3</sub>-32  $\delta$  0.96) and downfield shifts for C-8 ( $\delta$  30.32; C-17  $\delta$  26.62), C-9 ( $\delta$  35.49; C-16  $\delta$  33.06), and H-11 ( $\delta$  1.53; H-14  $\delta$  0.80).

TABLE 1.  $^{13}\text{C}$ -Nmr Chemical Shifts of Raspacionin [1] and Related Triterpenes 5-12.<sup>a</sup>

Carbon	1	5	6	7	8	9	10	11	12
C-1	43.37 <sup>b</sup>	43.44	42.82*	42.16	42.14	42.14	42.85	42.14	42.03
C-2	33.90	33.85	34.24**	40.27	40.14	40.27	35.89*	40.30*	41.32*
C-3	26.10	26.05	25.32	35.07	35.02	35.06	23.20**	34.98	35.03**
C-4	77.00	77.06	76.96 <sup>c</sup>	217.51	217.56	217.30	78.76	nd <sup>d</sup>	217.78 <sup>e</sup>
C-5	77.57	77.56	77.88	82.66	82.65	82.64	77.47	82.68	82.63
C-7	76.01	76.03	75.84	80.70	80.68	80.63	76.16	80.60	80.97
C-8	33.02	33.09	30.32	30.24	30.37	30.39	30.50	30.35	26.45
C-9	35.74	35.76	35.49	35.26	35.24	35.29	35.38	35.25	40.00
C-10	147.17	147.28	87.13	86.40	86.27	86.29	86.92	86.29	73.22
C-11	53.65	54.00	56.06	55.65	55.68	55.87	56.19	55.58	57.93
C-12	27.59	27.97	28.17	27.97	27.96	27.95	28.00	27.94	28.76
C-13	25.48	25.88	29.10	29.02	28.87	29.02	29.05 <sup>f</sup>	28.83	29.79
C-14	58.18	58.08	57.99	57.95	55.68	55.87	56.02	55.52	57.93
C-15	83.81	83.92	84.25	84.11	72.20	72.08	72.25	72.00	83.58
C-16	33.21	33.09	33.06	33.03	39.25	39.37	39.41	39.27	32.91
C-17	26.59	26.73	26.62	26.60	26.54	26.43	26.43	26.40	30.12
C-18	76.74	76.29	76.21	76.10	76.25	76.62	76.64	81.03	80.97
C-20	77.33	77.87	77.88	77.91	77.76	nd <sup>d</sup>	77.47	82.42	82.57
C-21	79.14	77.06	77.03 <sup>g</sup>	76.96	77.17	79.07	79.02	nd <sup>d</sup>	217.55 <sup>e</sup>
C-22	23.08	25.34	25.32	25.25	25.09	23.02	23.10**	34.98	35.09**
C-23	35.32	34.19	34.63**	34.32	34.12	35.42	35.49*	39.88*	40.49*
C-24	42.80 <sup>b</sup>	43.06	42.96*	42.81	42.55	42.45	42.45	41.84	41.75
C-25	12.20	12.13	13.16	12.53	12.49	12.65	13.27	12.50	12.25
C-26	29.11	29.12	28.90 <sup>oo</sup>	26.45	26.42	26.43	29.00 <sup>o</sup>	26.57	26.48
C-27	21.23	21.23	21.46	20.48	20.45	20.46	21.53 <sup>oo</sup>	20.46	20.46
C-28	107.38	107.36	19.71	19.85	19.69	19.71	19.72	19.77	23.54
C-29	25.26	25.25	24.67	24.73	30.31	30.39	30.41	30.52	24.48
C-30	21.56	21.46	21.46	21.43	21.31	21.49	21.48 <sup>oo</sup>	20.47	20.46
C-31	28.96	29.04	29.00 <sup>oo</sup>	28.96	29.00	28.96	28.81 <sup>o</sup>	26.40	26.45
C-32	12.81	12.81	12.92	12.85	12.93	12.90	13.09	12.20	12.25
OCOCH <sub>3</sub> -4	—	—	—	—	—	—	170.23	—	—
OCOCH <sub>3</sub> -10	—	—	170.09	170.06	170.20	170.12	170.23	nd <sup>d</sup>	—
OCOCH <sub>3</sub> -15	170.16	170.29	169.93	170.00	—	—	—	—	170.09
OCOCH <sub>3</sub> -21	170.16	—	—	—	—	170.12	170.23	—	—
OCOCH <sub>3</sub> -4	—	—	—	—	—	—	21.29	—	—
OCOCH <sub>3</sub> -10	—	—	22.88	22.82	23.05	23.08	23.20	23.07	—
OCOCH <sub>3</sub> -15	22.51	22.52	22.40	22.38	—	—	—	—	22.53
OCOCH <sub>3</sub> -21	21.23	—	—	—	—	21.28	21.29	—	—

<sup>a</sup>CDCl<sub>3</sub>; Bruker AMX-500 spectrometer. Chemical shifts referenced to CDCl<sub>3</sub> at 77.00 ppm.<sup>b</sup>These values were erroneously reported (1) inverted.<sup>d</sup>Nd=Not detected.\*\*\*<sup>oo</sup>Values with the same superscripts may be interchanged.

Surprisingly, the resonance of C-13 ( $\delta$  29.10) was shifted downfield almost 4 ppm from the values recorded for **1** ( $\delta$  25.48) and **5** ( $\delta$  25.88). It is likely that a dominant conformation displaying a  $\delta$  effect between C-13 and the substituent at C-10 can explain this apparent anomaly. All the  $^1\text{H}$ - and  $^{13}\text{C}$ -nmr resonance assignments were confirmed by 2D experiments (Tables 1 and 2).

10-Acetoxy-21-deacetyl-4-oxo-28-hydroraspacionin [**7**] exhibited an elemental composition of  $\text{C}_{34}\text{H}_{56}\text{O}_8$ , based on hreims of the ion fragment peak at  $m/z$  514.3645 [ $\text{M}-\text{HOAc}-\text{H}_2\text{O}$ ]<sup>+</sup>; 514.3658 calcd for  $\text{C}_{32}\text{H}_{50}\text{O}_5$ . The spectral data of **7** were compared with those of **6** revealing an additional  $^{13}\text{C}$ -nmr resonance at  $\delta$  217.51 that, along with the absence of one of the two carbinol protons at  $\delta$  3.82, supported the presence in **7** of a carbonyl group at C-4. Moreover, comparison with **6** revealed that the different functionalization at C-4 induced some diagnostic shifts for all protons and carbons of the seven-membered ring and, in addition, H-7 and H-11 were shifted upfield to  $\delta$  2.97 ( $\delta$  3.63 for **6**) and  $\delta$  1.46 ( $\delta$  1.53 for **6**), respectively. All  $^1\text{H}$ - and  $^{13}\text{C}$ -nmr resonances were confirmed by 2D experiments (Tables 1 and 2).

10-Acetoxy-15,21-dideacetyl-4-oxo-28-hydroraspacionin [**8**] possessed the molecular formula  $\text{C}_{32}\text{H}_{54}\text{O}_7$ , deduced by hreims at  $m/z$  472.3535 [ $\text{M}-\text{HOAc}-\text{H}_2\text{O}$ ]<sup>+</sup>,

TABLE 2. <sup>1</sup>H-Nmr Chemical Shifts of Raspacionin [1] and of the Triterpenoids 5–12.<sup>a</sup>

Proton	1	5	6	7	8	9	10	11	12
H-2	1.48	1.49	1.59	1.32	1.33	1.32	1.26	1.31	1.29
	1.65	1.66	nd <sup>b</sup>	1.87	1.87	1.80	1.41	1.80	1.79
H-3	1.82	1.79	1.74	2.13	2.13	2.13	1.75	2.13	2.14*
	2.00	1.98	2.02	3.18	3.18	3.20	2.00	3.19	3.20
H-4	3.83	3.82	3.82	—	—	—	4.99	—	—
H-7	3.68	3.67	3.63	2.97	2.97	2.96	3.48	2.95	2.92**
H-8	1.38	1.36	1.43	1.33	1.36	1.27	1.41	1.30	nd <sup>b</sup>
	1.62	1.63	1.53	1.61	1.56	1.58	1.57	1.61	nd <sup>b</sup>
H-9	1.95	1.95	1.66	1.67	1.69	1.62	1.66	1.67	1.79
	2.24	2.25	2.63	2.65	2.67	2.68	2.66	2.66	nd <sup>b</sup>
H-11	1.64	1.66	1.53	1.46	1.45	1.42	1.47	1.43	1.00
H-12	1.50	1.52	1.47	nd <sup>b</sup>	nd <sup>b</sup>	nd <sup>b</sup>	1.34*	nd <sup>b</sup>	1.29
	1.62	1.68	nd <sup>b</sup>	1.60	1.62	1.60	1.57	1.60	1.54
H-13	1.18	1.12	nd <sup>b</sup>	1.35	1.30	1.33	1.38*	nd <sup>b</sup>	1.29
	1.72	1.68	1.77	1.74	1.69	1.70	1.75	1.74	1.75
H-14	0.74	0.83	0.80	0.79	0.83	0.74	0.76	0.69	0.66
H-16	1.31	1.26	1.26	1.26	1.47	1.45	1.47	1.45	1.23
	2.80	2.76	2.78	2.78	1.60	1.61	1.64	1.64	2.84
H-17	1.39	1.36	1.36	1.37	1.39	1.40	1.41	1.45	1.47
	1.51	1.43	1.46	1.46	1.71	1.74	1.73	1.88	1.60
H-18	3.41	3.55	3.56	3.55	3.50	3.35	3.37	2.89	2.93**
H-21	4.97	3.82	3.82	3.83	3.82	4.97	4.98	—	—
H-22	1.81	1.75	1.74	1.74	1.73	1.76	1.75	2.09	2.12*
	2.00	1.98	2.02	2.03	2.02	1.98	2.00	3.23	3.23
H-23	1.27	1.38	1.45	1.38	1.37	1.22	1.26	1.17	1.20
	1.45	1.57	1.69	1.56	1.53	1.60	1.41	1.80	1.79
H <sub>3</sub> -25	0.70	0.69	0.83	0.98	0.99	0.99	0.87	0.96	0.93
H <sub>3</sub> -26	1.12	1.12*	1.11*	1.25	1.25	1.25	1.20**	1.27*	1.27 <sup>o</sup>
H <sub>3</sub> -27	1.27	1.26**	1.26**	1.31	1.31	1.31	1.14 <sup>o</sup>	1.30**	1.31 <sup>oo</sup>
H's-28	4.60	4.61	1.48	1.50	1.50	1.50	1.50	1.50	1.18
	4.89	4.88	—	—	—	—	—	—	—
H <sub>3</sub> -29	1.53	1.51	1.53	1.52	1.17	1.20	1.24	1.18	1.53
H <sub>3</sub> -30	1.15	1.27**	1.27**	1.26	1.27	1.15	1.15 <sup>o</sup>	1.31**	1.30 <sup>oo</sup>
H <sub>3</sub> -31	1.20	1.11*	1.13*	1.14	1.13	1.20	1.18**	1.25*	1.26 <sup>o</sup>
H <sub>3</sub> -32	0.94	0.93	0.96	0.94	0.99	0.97	1.00	1.12	1.08
OCOCH <sub>3</sub> -4	—	—	—	—	—	—	2.15 <sup>oo</sup>	—	—
OCOCH <sub>3</sub> -10	—	—	1.89	1.90	1.97	1.96	1.98	1.96	—
OCOCH <sub>3</sub> -15	1.95	1.93	1.93	1.93	—	—	—	—	1.96
OCOCH <sub>3</sub> -21	2.17	—	—	—	—	2.15	2.16 <sup>oo</sup>	—	—

<sup>a</sup>CDCl<sub>3</sub>; Bruker AMX-500 spectrometer. Chemical shifts referenced to CHCl<sub>3</sub> at 7.26 ppm.<sup>b</sup>Nd=Not detected.\*\*\*,<sup>o</sup>,<sup>oo</sup>Values with the same superscripts may be interchanged.

(C<sub>30</sub>H<sub>48</sub>O<sub>4</sub> requires 472.3552) and showed a <sup>13</sup>C-nmr signal at δ 217.56 that suggested the presence of a C=O group; its <sup>1</sup>H- and <sup>13</sup>C-nmr spectra are reported in Tables 1 and 2. The <sup>1</sup>H-nmr spectrum of **8** was almost identical to that of **7** with the only differences due to the absence of the acetyl group at C-15 that shifted C-15 to δ 72.20 (δ 84.11 for **7**), C-16 to δ 39.25 (δ 33.03 for **7**), C-29 to δ 30.31 (δ 24.73 for **7**) and, surprisingly, C-14 to δ 55.68 (δ 57.95 for **7**). All the nmr resonances were confirmed by 2D experiments. In particular, HMBC experiments exhibited a series of <sup>1</sup>H-<sup>13</sup>C long-range hetero-correlations (see Experimental) that further supported the assignments of all other raspacionin derivatives.

10-Acetoxy-15-deacetyl-4-oxo-28-hydroraspacionin [**9**] gave C<sub>34</sub>H<sub>56</sub>O<sub>8</sub>, assigned by hreims at *m/z* 532.3752 [M-HOAc]<sup>+</sup> (C<sub>32</sub>H<sub>52</sub>O<sub>6</sub> requires 532.3764). The spectral data were closely related to those of **8** and, in particular, the <sup>1</sup>H-nmr resonance at δ 4.97 (H-21; δ 3.82 for **8**), suggested a structure **9** that was easily confirmed by acetylation of **8**.

10-Acetoxy-4-acetyl-15-deacetyl-28-hydroraspacionin [**10**] exhibited C<sub>36</sub>H<sub>60</sub>O<sub>9</sub>, that was assigned by hreims at *m/z* 516.3803 [M-2 HOAc]<sup>+</sup> (C<sub>32</sub>H<sub>52</sub>O<sub>5</sub> requires 516.3814). The <sup>1</sup>H- and <sup>13</sup>C-nmr spectra of **10** (Tables 1 and 2) displayed resonances similar to those of **9** but with the <sup>13</sup>C-nmr resonance at δ 217.30 substituted by a signal

at  $\delta$  78.76, consistent with the presence of a secondary carbon (C-4) bearing an acetoxy group. The  $\beta$ -orientation of the substituent at C-4 was suggested by comparison of the nmr data of **10** (C-4  $\delta$  78.76, H-4  $\delta$  4.99) with those, already reported (**2**), of **2** (C-21  $\delta$  80.80, H-21  $\delta$  4.70). Compound **10** is also related to **6**, but with a different acetylation pattern. In fact, the  $^1\text{H}$ -nmr resonances of H-4 and H-21 were shifted to  $\delta$  4.99 and 4.98, respectively, whereas the deacetylation at C-15 was supported by the diagnostic  $^{13}\text{C}$ -nmr values of C-15 ( $\delta$  72.25), C-16 ( $\delta$  39.41) and C-29 ( $\delta$  30.41).

10-Acetoxy-15-deacetyl-4,21-dioxo-28-hydroraspacionin [**11**] possessed the molecular formula  $\text{C}_{32}\text{H}_{52}\text{O}_7$ , as determined by hreims at  $m/z$  488.3490 [ $\text{M}-\text{HOAc}$ ] $^+$  ( $\text{C}_{30}\text{H}_{48}\text{O}_5$ , requires 488.3501). The nmr data of **11** suggested a structure closely related to **10** but displaying two carbonyls at C-4 and C-21. It differed from **9** only by the functionalization at C-21 that shifted  $\text{H}_3-32$  to  $\delta$  1.12 (0.97 for **9**) and H-18 to  $\delta$  2.89 (3.35 for **9**). The perhydrobenzoxepine half of **11**, bearing the hydroxy group, displayed  $^1\text{H}$ - and  $^{13}\text{C}$ -nmr values almost identical to those of the corresponding partial structure in siphonone A [**13**] (**12**).

10-Hydroxy-4,21-dioxo-28-hydroraspacionin [**12**] exhibited  $\text{C}_{32}\text{H}_{52}\text{O}_7$  by hreims at  $m/z$  488.3482 [ $\text{M}-\text{HOAc}$ ] $^+$  ( $\text{C}_{30}\text{H}_{48}\text{O}_5$ , requires 488.3501). The same functionalization of **11** was observed in **12**, but with interchange of the acetoxy and hydroxy groups at carbons 10 and 15. The different functionalization of the oxygenated substituents at C-10 and C-15 induced a series of shifts on the vicinal carbons, including a surprising  $^{13}\text{C}$ -nmr downfield acetylation effect on C-14 ( $\delta$  57.93 for **12**;  $\delta$  55.52 for **11**). By analogy with **11**, the unacetylated unit of **12** was identical to the perhydrobenzoxepine moiety of sodwanone A [**14**] (**16**) as confirmed by comparison of nmr data.

All raspacionins (**7**, **8**, **9**, **11**, and **12**) containing a ketone moiety displayed a positive cd maximum at ca. 302 nm that suggested an absolute stereochemistry identical to those of the 4-oxo-derivative of raspacionin [**1**], the 21-oxo-derivative of raspacionin A [**2**], and raspacionin B [**3**] (**3**). The same absolute stereochemistry is presumably also possessed by the other raspacionins.

## EXPERIMENTAL

GENERAL EXPERIMENTAL PROCEDURES.—Ft-ir spectra were recorded with a Biorad FTS-7 instrument. Low-resolution ms were determined on a VG Trio-2000. High-resolution ms were determined on a Kratos MS-50 spectrometer. Optical rotations were recorded on a Jasco DIP-370 polarimeter. Cd measurements were carried out on a Jasco J-710 dicograph. Hplc was performed on a Waters apparatus equipped with a differential refractometer. Commercial Si gel (70–230 mesh ASTM) was used for column chromatography. Analytical tlc was carried out using precoated Si gel Merck  $\text{F}_{254}$  plates.

1D and 2D nmr spectra were recorded at room temperature with a Bruker AMX-500 spectrometer ( $^1\text{H}$ , 500.13 MHz;  $^{13}\text{C}$ , 125.76 MHz), equipped with a X32 data system.  $^1\text{H}$ - and  $^{13}\text{C}$ -nmr chemical shifts were referenced to  $\text{CHCl}_3$ , resonating at  $\delta$  7.26 and 77.00 ppm, respectively. The DEPT spectra were obtained using polarization transfer pulses of  $135^\circ$ . Two dimensional experiments were performed using standard Bruker microprograms.

EXTRACTION AND ISOLATION OF RASPACIONINS.—The sponge *Raspaciona aculeata* was collected by hand in Blanes (northeastern Spain) during December 1991, using scuba in an overhang at a depth of 10–15 m. A voucher specimen is deposited at the Centre d'Estudios Avanzados (Blanes). The  $\text{Et}_2\text{O}$ -soluble fraction (1.15 g) from the  $\text{Me}_2\text{CO}$  extract of the fresh sponge (dry wt 12.0 g) was fractionated on a Si gel flash column using light petroleum ether with increasing amounts of  $\text{Et}_2\text{O}$  as eluent to afford, along with the main metabolite **1** (158 mg) (tlc,  $R_f$  0.45, light petroleum ether- $\text{Et}_2\text{O}$ , 1:1) and a fraction (115 mg) containing a mixture of **2** and **3** (tlc,  $R_f$  0.70, light petroleum ether- $\text{Et}_2\text{O}$ , 1:1), a more polar fraction (116 mg) (tlc,  $R_f$  0.27, light petroleum ether- $\text{Et}_2\text{O}$ , 3:7) containing a mixture of the new triterpenoids **5**–**12**. This fraction was chromatographed by hplc using a Spherisorb Silica S5W column (25 cm  $\times$  10 mm, particle size 5  $\mu\text{m}$ , flow rate 2.5 ml/min $^{-1}$ ) and *n*-hexane- $\text{EtOAc}$  (7:3) as eluent, yielding seven main fractions: A (4.8 mg), B (17.0 mg), C (11.0 mg), D (6.0 mg), E (13.0 mg), F (10.0 mg), and G (6.8 mg). Each fraction was further purified by hplc using a Spherisorb 5 Sil column (25 cm  $\times$  4.6 mm, particle size 5  $\mu\text{m}$ ) and different eluents. In particular, elution of A with  $\text{CHCl}_3$  (flow rate 1.0 ml/min $^{-1}$ ) yielded 3.1 mg of **11**; elution of B with *n*-

hexane-*i*-PrOH (92:8) (flow rate 1.0 ml/min<sup>-1</sup>) yielded 5.7 mg of **9** and 3.5 mg of **7**; elution of C with *n*-hexane-*i*-PrOH (92:8) (flow rate 1.0 ml/min<sup>-1</sup>) yielded 2.7 mg of **5**; elution of D with CHCl<sub>3</sub> (flow rate 1.0 ml/min<sup>-1</sup>) yielded 3.1 mg of **12**; elution of E with CHCl<sub>3</sub> (flow rate 1.0 ml/min<sup>-1</sup>) yielded 2.5 mg of **10**; elution of F with *n*-hexane-*i*-PrOH (9:1) (flow rate 1.5 ml/min<sup>-1</sup>) yielded 2.6 mg of **6**; elution of G with *n*-hexane-*i*-PrOH (9:1) (flow rate 1.5 ml/min<sup>-1</sup>) yielded 2.7 mg of **8**.

**METHANOLYSIS OF 1 TO GIVE 5.**—Raspacionin [**1**] (2.5 mg) was treated with 1.0 ml of a solution of KOH (3% in MeOH) stirring at room temperature. The reaction was monitored by tlc (light petroleum ether-Et<sub>2</sub>O, 4:6) and stopped after 2 h, when the starting product spot disappeared. The usual work up gave 3.0 mg of a residue that was purified on Si gel contained in a Pasteur pipette using petroleum ether-Et<sub>2</sub>O (3:7) as eluent, to give 2.0 mg of pure compound **5**.

**ACETYLATION OF 8 TO GIVE 9.**—A few drops of Ac<sub>2</sub>O were added to a solution of **8** (1.4 mg) in dry pyridine (500 ml) and the reaction mixture was kept at room temperature overnight. After removal of the solvent *in vacuo*, the usual work up gave 1.0 mg of pure compound **9**.

**Raspacionin [1].**—Cd ( $c = 5.78 \times 10^{-4}$  M; EtOH) 20° [ $\theta$ ]<sub>206.60</sub> + 3469.

**21-Deacetyl-raspacionin [5].**—Amorphous powder: [ $\alpha$ ]<sub>D</sub><sup>25</sup> -49.1° ( $c = 0.25$ , CHCl<sub>3</sub>); cd ( $c = 5.20 \times 10^{-4}$  M; EtOH) 20° [ $\theta$ ]<sub>206.50</sub> + 2677; ir  $\nu$  max (liquid film, CHCl<sub>3</sub>) 2974, 2934, 1730, 1712 cm<sup>-1</sup>; eims *m/z* 474 [M-C<sub>2</sub>H<sub>4</sub>O<sub>2</sub>]<sup>+</sup> (5), 416 [M-C<sub>2</sub>H<sub>4</sub>O<sub>2</sub>-C<sub>3</sub>H<sub>6</sub>O]<sup>+</sup> (5), 398 [M-C<sub>2</sub>H<sub>4</sub>O<sub>2</sub>-C<sub>3</sub>H<sub>6</sub>O-H<sub>2</sub>O]<sup>+</sup> (2), 372 (10), 314 (15); hreims *m/z* 474.3698 [M-C<sub>2</sub>H<sub>4</sub>O<sub>2</sub>]<sup>+</sup> (C<sub>30</sub>H<sub>50</sub>O<sub>4</sub> requires 474.3709); <sup>1</sup>H- and <sup>13</sup>C-nmr data, see Tables 1 and 2.

**10-Acetoxy-21-deacetyl-28-hydroraspacionin [6].**—Amorphous powder: [ $\alpha$ ]<sub>D</sub><sup>25</sup> -27.7° ( $c = 0.26$ , CHCl<sub>3</sub>); cd ( $c = 4.86 \times 10^{-4}$  M; EtOH) 20° [ $\theta$ ]<sub>210.60</sub> -80.76; ir  $\nu$  max (liquid film, CHCl<sub>3</sub>) 2973, 2933, 2866, 1727 cm<sup>-1</sup>; eims *m/z* 474 [M-2C<sub>2</sub>H<sub>4</sub>O<sub>2</sub>]<sup>+</sup> (3), 416 [M-2C<sub>2</sub>H<sub>4</sub>O<sub>2</sub>-C<sub>3</sub>H<sub>6</sub>O]<sup>+</sup> (5), 314 (7); hreims *m/z* 474.3684 (C<sub>30</sub>H<sub>50</sub>O<sub>4</sub> requires 474.3709); <sup>1</sup>H- and <sup>13</sup>C-nmr data, see Tables 1 and 2.

**10-Acetoxy-21-deacetyl-4-oxo-28-hydroraspacionin [7].**—Amorphous powder: [ $\alpha$ ]<sub>D</sub><sup>25</sup> -38.5° ( $c = 0.37$ , CHCl<sub>3</sub>); cd ( $c = 4.17 \times 10^{-4}$  M; EtOH) 20° [ $\theta$ ]<sub>209.50</sub> + 9472, [ $\theta$ ]<sub>302.50</sub> + 8897; ir  $\nu$  max (liquid film, CHCl<sub>3</sub>) 2972, 2940, 2867, 1718 cm<sup>-1</sup>; eims *m/z* 514 [M-C<sub>2</sub>H<sub>4</sub>O<sub>2</sub>-H<sub>2</sub>O]<sup>+</sup> (0.15), 472 [M-2C<sub>2</sub>H<sub>4</sub>O<sub>2</sub>]<sup>+</sup> (13), 414 [M-2C<sub>2</sub>H<sub>4</sub>O<sub>2</sub>-C<sub>3</sub>H<sub>6</sub>O]<sup>+</sup> (4), 386 (2); hreims *m/z* 514.3645 [M-C<sub>2</sub>H<sub>4</sub>O<sub>2</sub>-H<sub>2</sub>O]<sup>+</sup> (C<sub>32</sub>H<sub>50</sub>O<sub>5</sub> requires 514.3658); <sup>1</sup>H- and <sup>13</sup>C-nmr data, see Tables 1 and 2.

**10-Acetoxy-15,21-dideacetyl-4-oxo-28-hydroraspacionin [8].**—Amorphous powder: [ $\alpha$ ]<sub>D</sub><sup>25</sup> -9.5° ( $c = 0.36$ , CHCl<sub>3</sub>); cd ( $c = 7.27 \times 10^{-4}$  M; EtOH) 20° [ $\theta$ ]<sub>209.90</sub> + 1200, [ $\theta$ ]<sub>302.10</sub> + 1109; ir  $\nu$  max (liquid film, CHCl<sub>3</sub>) 2922, 2852, 1716 cm<sup>-1</sup>; eims *m/z* 472 [M-C<sub>2</sub>H<sub>4</sub>O<sub>2</sub>-H<sub>2</sub>O]<sup>+</sup> (1), 432 [M-C<sub>2</sub>H<sub>4</sub>O<sub>2</sub>-C<sub>3</sub>H<sub>6</sub>O]<sup>+</sup> (2), 414 [M-C<sub>2</sub>H<sub>4</sub>O<sub>2</sub>-C<sub>3</sub>H<sub>6</sub>O-H<sub>2</sub>O]<sup>+</sup> (1), 372 (3); hreims *m/z* 472.3535 (C<sub>30</sub>H<sub>48</sub>O<sub>4</sub> requires 472.3552); <sup>1</sup>H- and <sup>13</sup>C-nmr data, see Tables 1 and 2; HMBC data ( $J = 10$  Hz)  $\delta$  42.14 (C-1) 0.99 (H<sub>3</sub>-25), 2.13 (H-3); 40.14 (C-2) 0.99 (H<sub>3</sub>-25), 2.13 (H-3), 2.97 (H-7), 3.18 (H-3); 35.02 (C-3) 1.87 (H-2); 217.56 (C-4) 3.18 (H-3); 82.65 (C-5) 1.25 (H<sub>3</sub>-26), 1.31 (H<sub>3</sub>-27), 2.13 (H-3), 2.97 (H-7); 80.68 (C-7) 0.99 (H<sub>3</sub>-25), 1.87 (H-2), 2.67 (H-9); 35.24 (C-9) 1.50 (H<sub>3</sub>-28), 86.27 (C-10) 1.50 (H<sub>3</sub>-28), 2.67 (H-9); 55.68 (C-11) 0.99 (H<sub>3</sub>-25), 1.50 (H<sub>3</sub>-28), 1.69 (H-9 and/or H-13), 2.67 (H-9); 27.96 (C-12) 0.83 (H-14), 1.45 (H-11); 28.87 (C-13) 0.83 (H-14); 55.68 (C-14) 0.97 (H<sub>3</sub>-32), 1.17 (H<sub>3</sub>-29), 1.60 (H-16); 72.20, (C-15) 1.17 (H<sub>3</sub>-29), 1.60 (H-16), 1.69 (H-13); 39.25 (C-16) 1.17 (H<sub>3</sub>-29); 76.25 (C-18) 0.97 (H<sub>3</sub>-32), 1.39 (H-17), 1.60 (H-16); 77.76 (C-20) 1.13 (H<sub>3</sub>-31), 1.27 (H<sub>3</sub>-30), 3.50 (H-18); 77.17 (C-21) 1.13 (H<sub>3</sub>-31), 1.27 (H<sub>3</sub>-30), 1.73 (H-22); 25.09 (C-22) 1.53 (H-23), 3.82 (H-21); 34.12 (C-23) 0.83 (H-14), 0.97 (H<sub>3</sub>-32), 2.02 (H-22), 3.50 (H-18), 3.82 (H-21); 42.55 (C-24) 0.83 (H-14), 0.97 (H<sub>3</sub>-32), 1.69 (H-13); 12.49 (C-25) 2.97 (H-7); 26.42 (C-26) 1.31 (H<sub>3</sub>-27); 20.45 (C-27) 1.25 (H<sub>3</sub>-26); 30.31 (C-29) 0.83 (H-14); 21.31 (C-30) 1.13 (H<sub>3</sub>-31); 29.00 (C-31) 1.27 (H<sub>3</sub>-30), 3.82 (H-21); 12.93 (C-32) 0.83 (H-14), 1.37 (H-23), 3.50 (H-18).

**10-Acetoxy-15-deacetyl-4-oxo-28-hydroraspacionin [9].**—Amorphous powder: [ $\alpha$ ]<sub>D</sub><sup>25</sup> -18.2° ( $c = 0.50$ , CHCl<sub>3</sub>); cd ( $c = 5.63 \times 10^{-4}$  M; EtOH) 20° [ $\theta$ ]<sub>210.50</sub> + 9288, [ $\theta$ ]<sub>302.60</sub> + 8689; ir  $\nu$  max (liquid film, CHCl<sub>3</sub>) 2973, 2940, 2862, 1721 cm<sup>-1</sup>; eims *m/z* 532 [M-C<sub>2</sub>H<sub>4</sub>O<sub>2</sub>]<sup>+</sup> (1), 472 [M-2C<sub>2</sub>H<sub>4</sub>O<sub>2</sub>]<sup>+</sup> (2), 414 [M-2C<sub>2</sub>H<sub>4</sub>O<sub>2</sub>-C<sub>3</sub>H<sub>6</sub>O]<sup>+</sup> (1); hreims *m/z* 532.3752, (C<sub>32</sub>H<sub>52</sub>O<sub>6</sub> requires 532.3764); <sup>1</sup>H- and <sup>13</sup>C-nmr data, see Tables 1 and 2.

**10-Acetoxy-4-acetyl-15-deacetyl-28-hydroraspacionin [10].**—Amorphous powder: [ $\alpha$ ]<sub>D</sub><sup>25</sup> -35.1° ( $c = 0.05$ , CHCl<sub>3</sub>); cd ( $c = 4.36 \times 10^{-4}$  M; EtOH) 20° [ $\theta$ ]<sub>206.50</sub> + 1820; ir  $\nu$  max (liquid film, CHCl<sub>3</sub>) 2971, 2938, 2865, 1726 cm<sup>-1</sup>; eims *m/z* 516 [M-2C<sub>2</sub>H<sub>4</sub>O<sub>2</sub>]<sup>+</sup> (1), 498 [M-2C<sub>2</sub>H<sub>4</sub>O<sub>2</sub>-H<sub>2</sub>O]<sup>+</sup> (1), 432 (3), 414 (4), 372 (3); hreims *m/z* 516.3803 (C<sub>32</sub>H<sub>52</sub>O<sub>6</sub> requires 516.3814); <sup>1</sup>H- and <sup>13</sup>C-nmr data, see Tables 1 and 2.

**10-Acetoxy-15-deacetyl-4,21-dioxo-28-hydroraspacionin [11].**—Amorphous powder: [ $\alpha$ ]<sub>D</sub><sup>25</sup> -36.1° ( $c = 0.1$ , CHCl<sub>3</sub>); cd ( $c = 6.08 \times 10^{-4}$  M; EtOH) 20° [ $\theta$ ]<sub>211.40</sub> + 10420, [ $\theta$ ]<sub>301.70</sub> + 10210; ir  $\nu$  max (liquid film, CHCl<sub>3</sub>) 2971, 2937, 2869, 1704 cm<sup>-1</sup>; eims *m/z* 488 [M-C<sub>2</sub>H<sub>4</sub>O<sub>2</sub>]<sup>+</sup> (2), 430 [M-C<sub>2</sub>H<sub>4</sub>O<sub>2</sub>-C<sub>3</sub>H<sub>6</sub>O]<sup>+</sup> (2),

412  $[M-C_2H_4O_2-C_3H_6O-H_2O]^+$  (2); hreims  $m/z$  488.3490 ( $C_{30}H_{48}O_5$ , requires 488.3501);  $^1H$ - and  $^{13}C$ -nmr data, see Tables 1 and 2.

*10-Hydroxy-4,21-dioxo-28-bydroraspacionin* [12].—Amorphous powder:  $[\alpha]_D^{25} - 5.9^\circ$  ( $c=0.28$ ,  $CHCl_3$ );  $cd$  ( $c=6.81 \times 10^{-4}$  M; EtOH)  $20^\circ$   $[\theta]_{302.70} + 6500$ ,  $[\theta]_{302.70} + 6569$ ;  $ir \nu_{max}$  (liquid film,  $CHCl_3$ ) 2974, 2936, 2863, 1707  $cm^{-1}$ ; eims  $m/z$  488  $[M-C_2H_4O_2]^+$  (2), 470  $[M-C_2H_4O_2-H_2O]^+$  (2), 430  $[M-C_2H_4O_2-C_3H_6O]^+$  (2), 412  $[M-C_2H_4O_2-C_3H_6O-H_2O]^+$  (2); hreims  $m/z$  488.3482 ( $C_{30}H_{48}O_5$ , requires 488.3501);  $^1H$ - and  $^{13}C$ -nmr data, see Tables 1 and 2.

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